



Occupational Health, Safety &
Environmental Consultants

Hamilton College
Limited Indoor Environmental Quality Assessment Report
for the Glenview Apartments

February 2024

Limited Indoor Environmental Quality Assessment Report

Client Site:

Hamilton College
198 College Hill Road
Clinton, New York 13323

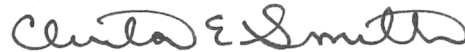
Prepared for:

Brian Hansen,
Director of Environmental Protection, Safety and Sustainability
Hamilton College

Colden Project #:


24123

Completed by:



Clinton E. Smith, MS, CIH, CSP
Industrial Hygienist and Project Manager

Reviewed by:



Shannon R. Magari, ScD, MS, MPH
Vice President, Health Sciences

Draft Report Date:

February 16, 2024

Final Report Date:

February 19, 2024

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EXECUTIVE SUMMARY

Hamilton College retained Colden Corporation to conduct a limited indoor environmental quality assessment in the Glenview Apartment housing units on campus in Clinton, New York. On January 16, 2024, Colden was contacted by Mr. Brian Hansen, the Director of Environmental Protection, Safety, and Sustainability for Hamilton College, regarding the presence of visible mold growth on surfaces inside dormitory rooms. Colden was asked to assist with identifying the type of mold growing, as well as investigate the potential source(s) and cause(s) of the growth as the focus of the initial site visit on January 19, 2024. Based on information provided prior to and during the on-site assessment, Colden completed the following scope of work in A125, an unoccupied room:

- Visual review of porous and non-porous surfaces,
- Visual review of the packaged terminal air conditioner (PTAC) unit,
- Visual review inside the wall cavity on two exterior walls and one interior wall using a borescope,
- Surface swab sampling for a fungal spore screen,
- Air sampling for a total fungal spore screen inside an interior wall cavity.

On January 29, 2024, Shannon R. Magari, ScD, MS, MPH, a Principal from Colden Corporation, toured the Glenview Apartments to observe conditions inside the individual rooms, lounges and bathrooms. On this day, Hamilton College asked Colden to perform air sampling in the Glenview Apartments the following day. The focus of the January 30, 2024, air sampling was to compare indoor air concentrations of total fungal spores inside the Glenview Apartments to outdoor concentrations. Based on information provided prior to and during the on-site assessment, Colden completed the following scope of work:

- Area air sampling for total fungal spores inside individual rooms, lounges, and outdoors for comparison purposes.

The on-site assessments were performed by Clinton Smith, MS, CIH, CSP, from Colden Corporation, a Mold Assessor licensed by the New York State Department of Labor. Brennan Borst of the Hamilton College EHS department was present and assisted during the assessments.

Results from the initial investigative efforts on January 19, 2024, confirmed the presence of contamination of *Aspergillus/Penicillium-like* mold spores on a wood stud inside the wall cavity of an interior wall between rooms A125 and A123. *Cladosporium* contamination was also confirmed on the surface of a PTAC fan blade in A125.

Based on these findings, Colden returned on January 30, 2024, to collect area air samples for total fungal spores in dorm rooms, lounges, and outdoors for comparison purposes. Average total fungal spore concentrations in all indoor samples were above outdoor concentrations except for B107.

1.0 INTRODUCTION

Hamilton College retained Colden Corporation to conduct a limited indoor environmental quality assessment in the Glenview Apartment housing units on campus in Clinton, New York. On January 16, 2024, Colden was contacted by Mr. Brian Hansen, the Director of Environmental Protection, Safety, and Sustainability for Hamilton College, regarding the presence of visible mold growth on surfaces inside dormitory rooms. Colden was asked to assist with identifying the type of mold growing, as well as investigate the potential source(s) and cause(s) of the growth.

1.1 Assessment Scope – January 19, 2024

The focus of the January 19, 2024, assessment was to identify the type of mold growing and investigate the potential source(s) and cause(s). Based on information provided prior to and during the on-site assessment, Colden completed the following scope of work in A125, an unoccupied room:

- Visual review of porous and non-porous surfaces,
- Visual review of the packaged terminal air conditioner (PTAC) unit,
- Visual review inside the wall cavity on two exterior walls and one interior wall using a borescope,
- Surface swab sampling for a fungal spore screen,
- Air sampling for a total fungal spore screen inside an interior wall cavity.

The on-site assessment was performed by Clinton Smith, MS, CIH, CSP, from Colden Corporation, a Mold Assessor licensed by the New York State Department of Labor. Brennan Borst of the Hamilton College EHS department was present and assisted during the assessment.

1.2 Assessment Scope – January 29 and 30, 2024

On January 29, 2024, Shannon R. Magari, ScD, MS, MPH, a Colden Principal, toured the Glenview Apartments to observe conditions inside the individual rooms, lounges and bathrooms. On this day Hamilton College asked Colden to perform air sampling in the Glenview Apartments the following day. The focus of the January 30, 2024, assessment was to compare indoor air concentrations of total fungal spores inside the Glenview Apartments to outdoor concentrations. Based on information provided prior to and during the on-site assessment, Colden completed the following scope of work:

- Area air sampling for total fungal spores in dorm rooms, lounges, and outdoors for comparison purposes.

2.0 METHODS

2.1 January 19, 2024

2.1.1 Visual Review

A visual review was performed in A125, an unoccupied room with two exterior walls and two interior walls. A representative from Hamilton College Facilities Management (FM) was present to cut three-inch holes through the 5/16-inch vinyl-over-gypsum (VOG) wall panels to expose sections of the wall cavity and interstitial space between rooms.

The front access panel of the PTAC unit and the metal grate shielding the fan were removed to allow for a visual review of the accessible interior surfaces of the unit.

2.1.2 Surface Swab Sampling

A total of eight swab samples were collected from the following surfaces to identify types of molds and qualitatively evaluate levels of contamination:

- Underside of mattress
- Interior surfaces of PTAC unit
- Surfaces of wood stud in wall cavity
- Backside of desk

Samples were collected using sterile swabs and analyzed by microscopy. The samples were analyzed by SGS Galson Laboratories of East Syracuse, New York, accredited by the American Industrial Hygiene Association (AIHA).

2.1.3 Air Sampling

As part of the investigative efforts to identify the potential source of mold amplification, air samples for total fungal spores were collected from within the interstitial space of the wall cavity using Zefon International Air-O-Cell™ cassettes and a Bio-Pump® calibrated to 15 liters per minute (LPM). The air samples were also submitted to SGS Galson for analysis.

2.2 January 30, 2024

2.2.1 Air Sampling

Area air samples for total fungal spores were collected in rooms throughout the A and B dorms on January 30, 2024. Six samples were collected in the A dorm and four samples were collected in the B dorm using Zefon International Air-O-Cell™ cassettes and a Bio-Pump® calibrated to 15 liters per minute (LPM). Air samples were also collected outdoors for comparison and reference purposes. The samples were analyzed by SGS Galson Laboratories. Two samples were collected at each location to account for the inherent variability in spore concentrations over time; the AIHA recommends at least two of the same type of air samples to assess variability of the data.

2.3 Guidelines for Indoor Airborne Mold

There are no published standards or guidelines for indoor concentrations of indoor airborne fungal spores. Competent authorities such as the AIHA recommend comparing indoor concentrations and genera with outdoor (or unaffected indoor areas) concentrations and genera to help determine whether mold is present at elevated concentrations and whether the space is an amplifier of mold.

3.0 RESULTS AND DISCUSSION

The results from the on-site assessment and sampling are summarized in the following sections. The laboratory analytical report and chain of custody are included in Attachment A.

3.1 January 19, 2024

Visible mold growth was present on the underside of both mattresses in A125. Visible growth was also present on the backside of one of the desks in the room. Surface wipe sampling results from samples HC240119-001 and HC240119-009, respectively, confirmed the “moderate” presence of *aspergillus/penicillium-like* spores on these two surfaces. Table 1 of this report summarizes results for all surface swab samples collected.

Interior surfaces of the PTAC unit had a visible accumulation of dust and other discolored material. *Aspergillus/Penicillium-like* was detected on all sampled surfaces of the PTAC unit. *Cladosporium* was detected on the surface of the fan blade (HC240119-003 & 004). Photos of the PTAC unit are provided for illustration as Photos 1 and 2 of the attached Photo Log.

Three-inch diameter holes were drilled behind the vinyl cove base on both exterior walls, and at one location on the interior wall. The visible building materials behind the holes on the two exterior walls did not appear to have visible microbial growth.

When the three-inch diameter VOG core was removed from the interior wall, as shown in Photo 3 of the Photo Log, cold air was felt exiting the hole into the room. Visible mold growth was present on wood studs inside this interior wall. Photos 4, 5, and 6 in the Photo Log, taken with a borescope, illustrate the presence of mold growth on the wood studs. Surface swab samples HC240119-006 & 007 confirmed the presence of *Aspergillus/Penicillium-like* spores. Air sampling results from within the interstitial space of the wall cavity also confirmed the presence of airborne *Aspergillus/Penicillium-like* spores.

3.2 January 30, 2024

The total fungal spore air sampling results are summarized in Table 2 and illustrated in Figure 1 of this report, comparing average indoor concentrations to average outdoor concentrations.

The average total fungal spore concentration outdoors in the parking lot near the A and B buildings was 165 spores per cubic meter of air (spores/m³). In the unoccupied rooms, average total fungal spore concentrations ranged from 230 to 10,900 spores/m³. In occupied dorm rooms, average total fungal spore concentrations ranged from 80 to 425 spores/m³. The average total fungal spore concentrations in the A and B lounges were 2,550 and 665 spores/m³, respectively.

4.0 CONCLUSIONS

Results from the initial investigative efforts on January 19, 2024, confirmed the presence of contamination of *Aspergillus/Penicillium-like* mold spores on a wood stud inside an interior wall between rooms A125 and A123. *Cladosporium* contamination was also confirmed on the surface of a PTAC fan blade in A125.

As a result of these findings, Colden returned on January 30, 2024, to collect area air samples for total fungal spores in dorm rooms, lounges, and outdoors for comparison purposes. Average total fungal spore concentrations in all indoor samples were above outdoor concentrations except for B107. There is amplification of mold on wood studs in the Glenview Apartments; additional investigation would be needed to determine the cause of the amplification.

5.0 LIMITATIONS

This report and its recommendations are based upon the conditions and observations during the time of the assessments. Reports do not purport to identify all hazards or unsafe practices, or to indicate that other hazards or unsafe practices do not exist. Colden does not assume and has no responsibility to the client for the control, correction, or continuance of conditions or practices, whether or not listed in this report, existing at the client's premises, or any other premises surveyed by Colden for and on behalf of the client. Colden's services shall be governed by the standard of practice for professional services as measured at the time those services are rendered.

Table 1
Surface Swab Fungal Spore Sampling Results from Glenview Apartment A125 – January 19, 2024

Sample ID	Sample Location	Photo No.	Level of Contamination ¹
HC240119-001	Underside of mattress	N/A	Aspergillus/Penicillium-like (Moderate)
HC240119-002	PTAC Unit – Rear internal surface of fan housing	5	Ascospores (Light) Aspergillus/Penicillium-like (Light) Cladosporium (Light) Basidiospores (Light)
HC240119-003	PTAC Unit – Surface of fan blade (right side)	5	Aspergillus/Penicillium-like (Light) Cladosporium (Heavy)
HC240119-004	PTAC Unit – Surface of fan blade (left side)	5	Ascospores (Light) Cladosporium (Heavy)
HC240119-005	PTAC Unit – Front internal surface of fan housing	4	Aspergillus/Penicillium-like (Light)
HC240119-006	Wall Cavity – Northeast side of wall stud	2	Aspergillus/Penicillium-like (Moderate)
HC240119-007	Wall Cavity – Southwest side of wall stud	6	Aspergillus/Penicillium-like (Heavy)
HC240119-009	Backside of desk on east side of room	N/A	Aspergillus/Penicillium-like (Moderate)

¹ According to SGS Galson, the level of contamination is a subjective measurement and corresponds to the general quantity of spores present in a sample. It also describes the amount of spores relative to one another.

NA – Not applicable; no photo.

Table 2
Summary of Average Airborne Total Fungal Spore Sampling Results – January 30, 2024

Sample IDs	Room Number	Sample Location/Description	Type of Mold Detected	Average Airborne Concentration (spores/m ³)
HC240130-A125A	A125	Unoccupied Dorm Room – Samples collected between beds. PTAC unit was not on during sample collection.	Aspergillus/Penicillium-like	150
HC240130-A125B			Basidiospores	40
			Cladosporium	40
			Total	230
			Aspergillus/Penicillium-like	150
			Basidiospores	67
			Cladosporium	13
			Total	230
HC240130-A126A	A126	Unoccupied Dorm Room – Samples collected between beds. PTAC unit was running during sample collection.	Aspergillus/Penicillium-like	5,700
HC240130-A126B			Basidiospores	53
			Cladosporium	40
			Total	5,793
			Aspergillus/Penicillium-like	7,500
			Basidiospores	27
			Cladosporium	0
			Total	7,527

Sample IDs	Room Number	Sample Location/Description	Type of Mold Detected	Average Airborne Concentration (spores/m ³)
HC240130-A120A	A120	Unoccupied Dorm Room – Samples collected between beds. PTAC unit was running during sample collection.	Ascospores Aspergillus/Penicillium-like Basidiospores Cladosporium Total	40 3,100 160 53 3,353
HC240130-A120B			Ascospores Aspergillus/Penicillium-like Basidiospores Cladosporium Total	150 9,200 160 890 10,400
HC240130-B125A	B125	Unoccupied Dorm Room – Samples collected between beds. PTAC unit was running during sample collection.	Aspergillus/Penicillium-like Basidiospores Cladosporium Total	16,000 13 0 16,013
HC240130-B125B			Aspergillus/Penicillium-like Basidiospores Cladosporium Total	1,600 0 27 1,627

Sample IDs	Room Number	Sample Location/Description	Type of Mold Detected	Average Airborne Concentration (spores/m ³)
HC240130-B109A	B109	Unoccupied Dorm Room – Samples collected between beds. PTAC unit was running during sample collection.	Aspergillus/Penicillium-like	8,500
			Basidiospores	80
			Cladosporium	67
			Total	8,647
HC240130-B109B			Aspergillus/Penicillium-like	13,000
			Basidiospores	40
			Cladosporium	170
			Total	13,210
HC240130-B107A	B107	Occupied Dorm Room – Samples collected between beds. PTAC unit was running during sample collection.	Aspergillus/Penicillium-like	0
			Basidiospores	13
			Cladosporium	0
			Total	13
HC240130-B107B			Aspergillus/Penicillium-like	120
			Basidiospores	27
			Cladosporium	0
			Total	147
HC240130-A105A	A105	Occupied Dorm Room – Samples collected between beds. PTAC unit was running during sample collection.	Ascospores	0
			Aspergillus/Penicillium-like	120
			Basidiospores	13
			Cladosporium	67
			Total	200
HC240130-A105B			Ascospores	27
			Aspergillus/Penicillium-like	110
			Basidiospores	27
			Cladosporium	13
			Total	177

Sample IDs	Room Number	Sample Location/Description	Type of Mold Detected	Average Airborne Concentration (spores/m ³)
HC240130-A118A	A118	Occupied Dorm Room – Samples collected between beds. PTAC unit was running during sample collection.	Aspergillus/Penicillium-like	810
HC240130-A118B			Basidiospores	13
			Cladosporium	0
			Total	823
			Aspergillus/Penicillium-like	13
			Basidiospores	13
			Cladosporium	0
			Total	26
HC240130-A119A	A119	Lounge – Samples collected near tv between couches. PTAC units were running during sample collection.	Ascospores	0
			Aspergillus/Penicillium-like	450
			Basidiospores	67
			Cladosporium	170
			Total	687
HC240130-A119B			Ascospores	27
			Aspergillus/Penicillium-like	2,700
			Basidiospores	280
			Cladosporium	1,400
			Total	4,407

Sample IDs	Room Number	Sample Location/Description	Type of Mold Detected	Average Airborne Concentration (spores/m ³)
HC240130-B119A	B119	Lounge – Samples collected near tv between couches. PTAC units were running during sample collection.	Aspergillus/Penicillium-like	240
HC240130-B119B			Basidiospores	290
			Cladosporium	0
			Total	530
			Aspergillus/Penicillium-like	590
			Basidiospores	170
			Cladosporium	40
			Total	800
HC240130-OUT1	Not Applicable	Outdoors – Samples collected in the parking lot near the 90° corner of the A and B buildings.	Ascospores	0
			Aspergillus/Penicillium-like	0
			Basidiospores	0
			Cladosporium	0
			Total	0
HC240130-OUT2			Ascospores	13
			Aspergillus/Penicillium-like	13
			Basidiospores	210
			Cladosporium	93
			Total	329

Spores/m³ – Average number of total fungal spores per cubic meter of air

Figure 1
Average Total Fungal Spore Air Sampling Results – January 30, 2024



UD – unoccupied dorm
OD – occupied dorm

ATTACHMENT A
Photo Log

Photo Number: 1

Location: A125

Description:

- PTAC unit with front access panel and fan grate removed.
- Sample HC240119-005 collected from front internal surface of fan housing with minimal visible dust and debris present.



Photo Number: 2

Location: A125

Description:

- Unknown discoloration on interior surface of PTAC fan housing. Surface sample HC240119-002.
- Visible mold growth on surface of PTAC fan blade. Surface samples HC240119-003 and 004.

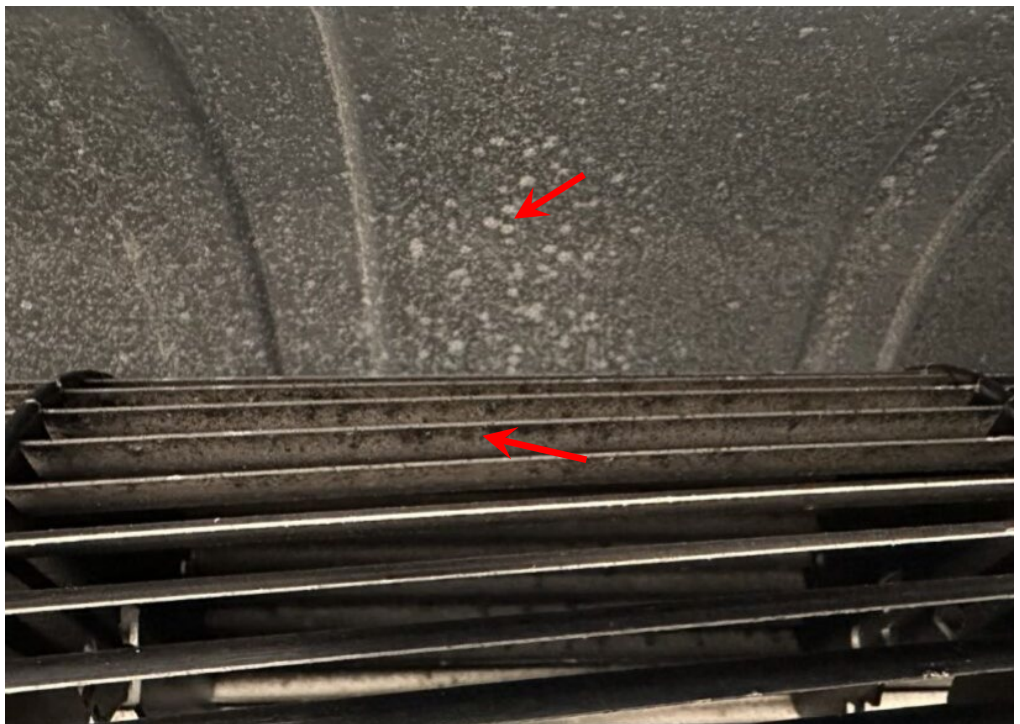


Photo Number: 3

Location: A125

Description:

- Three-inch holes cut through the interior wall.
- Air samples HC240119-008A and 008B were collected from inside the hole furthest from the floor.



Photo Number: 4

Location: A125

Description:

- Visible mold growth on wall stud of interior wall as seen with the borescope through three-inch hole.



Photo Number: 5

Location: A125

Description:

- Visible mold growth on wall stud of interior wall as seen with the borescope through three-inch hole.



Photo Number: 6

Location: A125

Description:

- Visible mold growth on wall stud of interior wall as seen with the borescope through three-inch hole.



ATTACHMENT B
Laboratory Reports and Chains of Custody



GALSON

**Clint Smith
Colden Corporation
5842 Heritage Landing Dr.
East Syracuse, NY 13057**

January 26, 2024

Account# 13111

Login# L615648

Dear Clint Smith:

Enclosed are the analytical results for the samples received by our laboratory on January 19, 2024. All samples on the chain of custody were received in good condition unless otherwise noted. Any additional observations will be noted on the chain of custody.

Please contact client services at (888) 432-5227 if you would like any additional information regarding this report. Thank you for using SGS Galson.

Sincerely,

SGS Galson

A handwritten signature in black ink that reads 'Lisa Swab'.

**Lisa Swab
Laboratory Director**

Enclosure(s)



Terms and Conditions & General Disclaimers

- This document is issued by the Company under its General Conditions of Service accessible at <http://www.sgs.com/en/Terms-and-Conditions.aspx>. Attention is drawn to the limitation of liability, indemnification and jurisdiction issues defined therein.
- Any holder of this document is advised that information contained herein reflects the Company’s findings at the time of its intervention only and within the limits of Client’s instructions, if any. The Company’s sole responsibility is to its Client and this document does not exonerate parties to a transaction from exercising all their rights and obligations under the transaction documents. Any unauthorized alteration, forgery or falsification of the content or appearance of this document is unlawful and offenders may be prosecuted to the fullest extent of the law.

Analytical Disclaimers

- Unless otherwise noted within the report, all quality control results associated with the samples were within established control limits or did not impact reported results.
- Note: The findings recorded within this report were drawn from analysis of the sample(s) provided to the laboratory by the Client (or a third party acting at the Client’s direction). The laboratory does not have control over the sampling process, including but not limited to the use of field equipment and collection media, as well as the sampling duration, collection volume or any other collection parameter used by the Client. The findings herein constitute no warranty of the sample's representativeness of any sampled environment, and strictly relate to the samples as they were presented to the laboratory. For recommended sampling collection parameters, please refer to the Sampling and Analysis Guide at www.sgsgalson.com.
- Unrounded results are carried through the calculations that yield the final result and the final result is rounded to the number of significant figures appropriate to the accuracy of the analytical method. Please note that results appearing in the columns preceding the final result column may have been rounded and therefore, if carried through the calculations, may not yield an identical final result to the one reported.
- The stated LOQs for each analyte represent the demonstrated LOQ concentrations prior to correction for desorption efficiency (if applicable).
- Unless otherwise noted within the report, results have not been blank corrected for any field blank or method blank data.

Accreditations SGS Galson holds a variety of accreditations and recognitions. Our quality management system conforms with the requirements of ISO/IEC 17025. Where applicable, samples may also be analyzed in accordance with the requirements of ELAP, NELAC, or LELAP under one of the state accrediting bodies listed below. Current Scopes of Accreditation can be viewed at <http://www.sgsgalson.com> in the accreditations section of the "About" page. To determine if the analyte tested falls under our scope of accreditation, please visit our website or call Client Services at (888) 432-5227.

National/International	Accreditation/Recognition	Lab ID#	Program/Sector
AIHA-LAP, LLC - IHLAP, ELLAP, EMLAP	ISO/IEC 17025 and USEPA NLLAP	Lab ID 100324	Industrial Hygiene, Environmental Lead, Environmental Microbiology

State	Accreditation/Recognition	Lab ID#	Program/Sector
New York (NYSDOH)	ELAP and NELAC (TNI)	Lab ID: 11626	Air Analysis, Solid and Hazardous Waste
Louisiana (LDEQ)	LELAP	Lab ID: 04083	Air Analysis, Solid Chemical Materials

Legend

< - Less than	mg - Milligrams	MDL - Method Detection Limit	ppb - Parts per Billion
> - Greater than	ug - Micrograms	NA - Not Applicable	ppm - Parts per Million
l - Liters	m3 - Cubic Meters	NS - Not Specified	ppbv - ppb Volume
LOQ - Limit of Quantitation	kg - Kilograms	ND - Not Detected	ppmv - ppm Volume
ft2 - Square Feet	cm2 - Square Centimeters	in2 - Square Inches	ng - Nanograms



GALSON

LABORATORY ANALYSIS REPORT

6601 Kirkville Road
 East Syracuse, NY 13057
 (315) 432-5227
 FAX: (315) 437-0571
 www.sgsgalson.com

Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 19-JAN-24
 Date Received : 19-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L615648
 Date Analyzed : 24-JAN-24
 Report ID : 1403510

Client ID : HC240119-008A Lab ID : L615648-8 Air Volume : 0.01498 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

WALL CAVITY

<u>Parameter</u>	<u>Raw Count</u>	<u>Total Count</u>	<u>Conc Count/m3</u>	<u>Percent %</u>
Mycelial Fragments	15	15	1000	NA
Pollen	<1	<1	<67	NA
Total Fungal Spores	481	725	48000	NA

<i>Alternaria</i>	<1	<1	<67	NA
Ascospores	6	6	400	0.8
<i>Aspergillus/Penicillium</i> -like	105	105	7000	14.5
Basidiospores	35	35	2300	4.8
<i>Bipolaris/Drechslera</i>	<1	<1	<67	NA
<i>Chaetomium</i>	<1	<1	<67	NA
<i>Cladosporium</i>	326	570	38000	78.6
<i>Curvularia</i>	<1	<1	<67	NA
Rusts/Smuts	<1	<1	<67	NA
<i>Stachybotrys</i>	<1	<1	<67	NA
Other/Unidentified	9	9	600	1.2

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: TAC Supervisor: BDB Date : 24-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : SLS Sampler : Spore Trap

CFU -Colony Forming Units g -Grams



GALSON

LABORATORY ANALYSIS REPORT

6601 Kirkville Road
East Syracuse, NY 13057
(315) 432-5227
FAX: (315) 437-0571
www.sgsgalson.com

Client : Colden Corporation
Site : HC
Project No. : 24123
Date Sampled : 19-JAN-24
Date Received : 19-JAN-24
Incubation Temp : NA

Account No.: 13111
Login No. : L615648
Date Analyzed : 24-JAN-24
Report ID : 1403510

Client ID : HC240119-008B Lab ID : L615648-9 Air Volume : 0.02996 m3
Analysis : Standard Mold Screen Crowding Factor : 1

WALL CAVITY

Table with 5 columns: Parameter, Raw Count, Total Count, Conc Count/m3, Percent %. Rows include Mycelial Fragments, Pollen, Total Fungal Spores, Alternaria, Ascospores, Aspergillus/Penicillium-like, Basidiospores, Bipolaris/Drechslera, Chaetomium, Cladosporium, Curvularia, Rusts/Smuts, Stachybotrys, and Other/Unidentified.

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: TAC Supervisor: BDB Date : 24-JAN-24
Analytical Method : In-house: IB-AIROCELL; Mic Approved by : SLS Sampler : Spore Trap

CFU -Colony Forming Units g -Grams



GALSON

LABORATORY ANALYSIS REPORT

6601 Kirkville Road
East Syracuse, NY 13057
(315) 432-5227
FAX: (315) 437-0571
www.sgsgalson.com

Client : Colden Corporation
Site : HC
Project No. : 24123
Date Sampled : 19-JAN-24
Date Received : 19-JAN-24
Incubation Temp : NA

Account No.: 13111
Login No. : L615648
Date Analyzed : 26-JAN-24
Report ID : 1403962

Client ID : HC240119-001 Lab ID : L615648-1 Swab Area : NA
Analysis : Standard Mold Screen

UNDERSIDE OF MATTRESS

<u>Parameter</u>	<u>Level of contamination</u>
Mycelial Fragments	Light
<i>Alternaria</i>	ND
Ascospores	ND
<i>Aspergillus/Penicillium</i> -like	Moderate
Basidiospores	ND
<i>Bipolaris/Drechslera</i>	ND
<i>Chaetomium</i>	ND
<i>Cladosporium</i>	ND
<i>Curvularia</i>	ND
Rusts/Smuts	ND
<i>Stachybotrys</i>	ND
Other/Unidentified	ND

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore	Submitted by: TAC	Supervisor: BDB	Date : 26-JAN-24
Analytical Method : In-house: MICR-SOP-21; Mic	Approved by : SLS		Sampler : Swab

CFU -Colony Forming Units g -Grams



GALSON

LABORATORY ANALYSIS REPORT

6601 Kirkville Road
East Syracuse, NY 13057
(315) 432-5227
FAX: (315) 437-0571
www.sgsgalson.com

Client : Colden Corporation
Site : HC
Project No. : 24123
Date Sampled : 19-JAN-24
Date Received : 19-JAN-24
Incubation Temp : NA

Account No.: 13111
Login No. : L615648
Date Analyzed : 26-JAN-24
Report ID : 1403962

Client ID : HC240119-002 Lab ID : L615648-2 Swab Area : NA
Analysis : Standard Mold Screen

**PTAC VENTILATION UNIT -
INTERNAL REAR OF HOUSING**

<u>Parameter</u>	<u>Level of contamination</u>
Mycelial Fragments	Light
<i>Alternaria</i>	ND
Ascospores	Light
<i>Aspergillus/Penicillium</i> -like	Light
Basidiospores	Light
<i>Bipolaris/Drechslera</i>	ND
<i>Chaetomium</i>	ND
<i>Cladosporium</i>	Light
<i>Curvularia</i>	ND
Rusts/Smuts	ND
<i>Stachybotrys</i>	ND
Other/Unidentified	Light

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore	Submitted by: TAC	Supervisor: BDB	Date : 26-JAN-24
Analytical Method : In-house: MICR-SOP-21; Mic	Approved by : SLS		Sampler : Swab

CFU -Colony Forming Units g -Grams



GALSON

LABORATORY ANALYSIS REPORT

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FAX: (315) 437-0571
www.sgsgalson.com

Client : Colden Corporation
Site : HC
Project No. : 24123
Date Sampled : 19-JAN-24
Date Received : 19-JAN-24
Incubation Temp : NA

Account No.: 13111
Login No. : L615648
Date Analyzed : 26-JAN-24
Report ID : 1403962

Client ID : HC240119-003 Lab ID : L615648-3 Swab Area : NA
Analysis : Standard Mold Screen

PTAC VENTILATION UNIT -
RIGHT SIDE OF FAN BLADE

<u>Parameter</u>	<u>Level of contamination</u>
Mycelial Fragments	Light
<i>Alternaria</i>	ND
Ascospores	ND
<i>Aspergillus/Penicillium</i> -like	Light
Basidiospores	ND
<i>Bipolaris/Drechslera</i>	ND
<i>Chaetomium</i>	ND
<i>Cladosporium</i>	Heavy
<i>Curvularia</i>	ND
Rusts/Smuts	ND
<i>Stachybotrys</i>	ND
Other/Unidentified	ND

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore	Submitted by: TAC	Supervisor: BDB	Date : 26-JAN-24
Analytical Method : In-house: MICR-SOP-21; Mic	Approved by : SLS		Sampler : Swab

CFU -Colony Forming Units g -Grams



GALSON

LABORATORY ANALYSIS REPORT

6601 Kirkville Road
East Syracuse, NY 13057
(315) 432-5227
FAX: (315) 437-0571
www.sgsgalson.com

Client : Colden Corporation
Site : HC
Project No. : 24123
Date Sampled : 19-JAN-24
Date Received : 19-JAN-24
Incubation Temp : NA

Account No.: 13111
Login No. : L615648
Date Analyzed : 26-JAN-24
Report ID : 1403962

Client ID : HC240119-004 Lab ID : L615648-4 Swab Area : NA
Analysis : Standard Mold Screen

**PTAC VENTILATION UNIT -
LEFT SIDE OF FAN BLADE**

<u>Parameter</u>	<u>Level of contamination</u>
Mycelial Fragments	Light
<i>Alternaria</i>	ND
Ascospores	Light
<i>Aspergillus/Penicillium</i> -like	ND
Basidiospores	ND
<i>Bipolaris/Drechslera</i>	ND
<i>Chaetomium</i>	ND
<i>Cladosporium</i>	Heavy
<i>Curvularia</i>	ND
Rusts/Smuts	ND
<i>Stachybotrys</i>	ND
Other/Unidentified	ND

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore	Submitted by: TAC	Supervisor: BDB	Date : 26-JAN-24
Analytical Method : In-house: MICR-SOP-21; Mic	Approved by : SLS		Sampler : Swab

CFU -Colony Forming Units g -Grams



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LABORATORY ANALYSIS REPORT

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East Syracuse, NY 13057
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FAX: (315) 437-0571
www.sgsgalson.com

Client : Colden Corporation
Site : HC
Project No. : 24123
Date Sampled : 19-JAN-24
Date Received : 19-JAN-24
Incubation Temp : NA

Account No.: 13111
Login No. : L615648
Date Analyzed : 26-JAN-24
Report ID : 1403962

Client ID : HC240119-005 Lab ID : L615648-5 Swab Area : NA
Analysis : Standard Mold Screen

**PTAC VENTILATION UNIT -
INTERNAL FRONT OF HOUSING**

<u>Parameter</u>	<u>Level of contamination</u>
Mycelial Fragments	Light
<i>Alternaria</i>	ND
Ascospores	ND
<i>Aspergillus/Penicillium</i> -like	Light
Basidiospores	ND
<i>Bipolaris/Drechslera</i>	ND
<i>Chaetomium</i>	ND
<i>Cladosporium</i>	ND
<i>Curvularia</i>	ND
Rusts/Smuts	ND
<i>Stachybotrys</i>	ND
Other/Unidentified	Light

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore	Submitted by: TAC	Supervisor: BDB	Date : 26-JAN-24
Analytical Method : In-house: MICR-SOP-21; Mic	Approved by : SLS		Sampler : Swab

CFU -Colony Forming Units g -Grams



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LABORATORY ANALYSIS REPORT

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 FAX: (315) 437-0571
 www.sgsgalson.com

Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 19-JAN-24
 Date Received : 19-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L615648
 Date Analyzed : 26-JAN-24
 Report ID : 1403962

Client ID : HC240119-006 Lab ID : L615648-6 Swab Area : NA
 Analysis : Standard Mold Screen

**WALL CAVITY -
 SURFACE OF WALL STUD**

<u>Parameter</u>	<u>Level of contamination</u>
Mycelial Fragments	Light
<i>Alternaria</i>	ND
Ascospores	ND
<i>Aspergillus/Penicillium</i> -like	Moderate
Basidiospores	ND
<i>Bipolaris/Drechslera</i>	ND
<i>Chaetomium</i>	ND
<i>Cladosporium</i>	ND
<i>Curvularia</i>	ND
Rusts/Smuts	ND
<i>Stachybotrys</i>	ND
Other/Unidentified	ND

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: TAC Supervisor: BDB Date : 26-JAN-24
 Analytical Method : In-house: MICR-SOP-21; Mic Approved by : SLS Sampler : Swab

CFU -Colony Forming Units g -Grams



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LABORATORY ANALYSIS REPORT

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 FAX: (315) 437-0571
 www.sgsgalson.com

Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 19-JAN-24
 Date Received : 19-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L615648
 Date Analyzed : 26-JAN-24
 Report ID : 1403962

Client ID : HC240119-007 Lab ID : L615648-7 Swab Area : NA
 Analysis : Standard Mold Screen

**WALL CAVITY -
 SURFACE OF WALL STUD**

<u>Parameter</u>	<u>Level of contamination</u>
Mycelial Fragments	Light
<i>Alternaria</i>	ND
Ascospores	ND
<i>Aspergillus/Penicillium</i> -like	Heavy
Basidiospores	ND
<i>Bipolaris/Drechslera</i>	ND
<i>Chaetomium</i>	ND
<i>Cladosporium</i>	ND
<i>Curvularia</i>	ND
Rusts/Smuts	ND
<i>Stachybotrys</i>	ND
Other/Unidentified	ND

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: TAC Supervisor: BDB Date : 26-JAN-24
 Analytical Method : In-house: MICR-SOP-21; Mic Approved by : SLS Sampler : Swab

CFU -Colony Forming Units g -Grams



GALSON

LABORATORY FOOTNOTE REPORT

6601 Kirkville Road
East Syracuse, NY 13057
(315) 432-5227
FAX: (315) 437-0571
www.ssggalson.com

Client Name : Colden Corporation
Site : HC
Project No. : 24123

Date Sampled : 19-JAN-24
Date Received: 19-JAN-24
Date Analyzed: 24-JAN-24 - 26-JAN-24

Account No.: 13111
Login No. : L615648

L615648 (Report ID: 1403962):
SOPs: MICR-SOP-21(3)

L615648 (Report ID: 1403510):
SOPs: ib-airocell(28)



6601 Kirkville Road
 East Syracuse, NY 13057-0369
 Phone: (888) 432-5227
 Fax: (315) 437-0571
<http://www.sgsgalson.com>

Analytical Notes for Microbiology Air-O-Cell™ Cassettes and other Spore Traps

Air-O-Cell™ cassettes and other spore traps may capture non-microbial particles that may interfere with spore counts. SGS Galson provides an estimation of the density of these particles, referred to as a Crowding Factor. The Crowding Factor ranges from 0 to 5 and is explained below. High levels of particulate matter on the impaction medium may bias the analysis by obscuring or covering spores. In addition, particle capture efficiency may decrease with high levels of particulate matter.

Crowding Factor	Explanation
0	No particles detected. This is typical of blank samples. Because most air samples typically contain some particles, absence of particulate matter could indicate improper sampling if the sample was not meant to be a blank.
1	Particles are far apart and in low numbers. Particulate matter covers approximately <5% of the impaction area. Spore counts not affected or minimally affected by the particle load.
2	Particles are close together and/or overlapping, and some spores may be obscured. Particulate matter covers approximately 5% to 25% of the impaction area. Spore counts may be biased low.
3	Particles are moderately crowded. It is likely that some spores are obscured. Particulate matter covers approximately 25% to 75% of the impaction area. Spore counts are likely biased low.
4	Particles are crowded, frequently obscuring spores. Particulate matter covers approximately 75% to 90% of the impaction area. Spore counts are likely biased low. The degree of bias increases with the percent of the trace that is occluded.
5	Particles are overcrowded making analysis impossible; no spore counts provided. If certain spores are readily detectable, they are reported as "Detected". If heavy quantities of spores are observed along the edges of the trace, this is footnoted in the report.



Counts for any genus that exceed 300 spores are estimated to two significant figures.

The list of fungal spores reported is:

***Alternaria* includes spores previously reported as *Ulocladium*.**

Ascospores – includes all ascospores with the exception of *Chaetomium*.

***Aspergillus/Penicillium*-like** – These two genera are grouped together as the spores are indistinguishable on a spore trap.

Basidiospores – This includes all basidiospores, even ones that can be identified to genus level, such as *Ganoderma*.

Bipolaris/Drechslera – *Helminthosporium* and *Exserohilum* are included in this grouping.

Chaetomium – Due to its unique shape and due to the fact that it may be associated with indoor mold problems, this ascospore is reported separate from other ascospores.

Cladosporium

Curvularia

Rusts/Smuts – *Myxomycetes* and *Periconia* are included in this grouping.

Stachybotrys – This includes *Memnoniella*.

Ulocladium* has been reclassified and is now reported as *Alternaria

Other/Unidentified – “Other” includes spores that can be identified but are rarely observed and/or are typically seen in small quantities. They include: *Acremonium*, *Botrytis*, *Cercospora*, *Epicoccum*, *Fusarium*, *Nigrospora*, *Oidium*, *Paecilomyces*, *Pestalotia*, *Pestalotiopsis*, *Pithomyces*, *Polythrincium*, *Scopulariopsis*, *Spegazzinia*, *Stemphylium*, *Taeniocella*, *Tetraploa*, *Torula*, and *Trichoderma*, and *Zygophiala*. “Unidentified” includes broken and dehydrated spores, spores that are partially obscured by debris, and spores that can't be categorized using microscopy alone.

In addition, other analytes that will be shown on reports include mycelial fragments (hyphae) and pollen.

Reports for expanded analysis include the above list with the addition of skin cells and fibers.

Generally, 100% of the sample deposit is analyzed. However, some analytes with high counts may be estimated based on the analysis of a portion of the slide and the results extrapolated. In these cases, the reported values will differ between the “Raw Count” and “Total Count” columns. For example, if an analyst observed 304 basidiospores after analyzing 25% of the sample, the estimated value is 1216. The final report would show 304 in the “Raw Count” column and 1200 in the “Total Count” column (the “Total Column” is rounded to two significant figures).



Direct Microscopic Examination (Screens)

- The analytes that we report are the same as those listed for spore traps with the exceptions of pollen, skin cells, and fibers.
- Due to the inherent nature of screen samples, a spore count is not performed.
- Upon special request counts may be performed on swab, liquid, or bulk screens. Counts are never performed on tape lifts due to the nature of the samples to not have uniform distribution of spores.
- The amount of a particular spore detected is reported as a “Level of contamination”. The level of contamination is a subjective measurement and corresponds to the general quantity of spores present in a sample. It also describes the amount of spores relative to one another.
 - Light: approximately 1 to 5 spores or mycelial fragments per microscope field of view at 600x.
 - Moderate: 6 to 15 spores or mycelial fragments per microscope field of view at 600x.
 - Heavy: Greater than 15 spores or mycelial fragments per microscope field of view at 600x.

Viable Fungi Analysis

- Standard growing conditions for viable fungi are $25^{\circ}\text{C} \pm 1^{\circ}\text{C}$ for 7 days.
- Standard growing conditions for viable thermophilic fungi are $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$ for 7 days.
- Results are reported in colony forming units (CFUs). A CFU can originate from one or many spores.
- SGS Galson uses and provides Potato Dextrose agar for all cultureable fungal methods. We have found Potato Dextrose agar to be suitable for the culture of the widest range of organisms. Other agars submitted or requested by clients are grown under the above standard conditions unless otherwise requested by the client.
- Some fungi may not produce identifiable structures in culture or under standard growing conditions. These fungi will be considered sterile hyphae and reported as such.
- Lack of growth under standard conditions does not preclude the presence of fungi or its viability in a sample.
- Samples taken with impactor samplers are not corrected for a positive hole correction factor.
- Identification of fungal organisms is based on visual microscopic examination at up to seven days of growth under standard conditions. Due to the large numbers of different species that may comprise them, certain genera may appear similar due to variations in stages of their life cycles, growth requirements, and/or environmental stress. A very limited amount of identification overlap may occur due to morphological similarities.
- Final interpretation of results is up to the person(s) responsible for conducting the sampling.



Quality Control/Quality Assurance

- A daily quality control spore trap slide is read each day that an analyst performs analysis on client spore trap samples. These slides consist of old client samples that have been analyzed a minimum of twenty times before they are used as a part of the quality control program. Control limits are set at the mean plus or minus three standard deviations for each analyte and for the total spore count. Warning limits are set at the mean plus or minus two standard deviations for each analyte and for the total spore count.
- A minimum of five percent of the samples are analyzed as duplicates and five percent of the samples are analyzed as replicates (or at least one replicate or duplicate per day). The relative percent difference (RPD) is calculated between the original sample result and its duplicate or replicate. The RPD value must fall within statistically based limits. In addition, there must be agreement between three of the top five categories.
- Daily quality control includes a blind spore trap challenge and a blind fungal culture identification challenge. Each analyst must correctly identify a spore or other airborne particulate from an old spore trap slide and identify a slide prepared from a fungal culture, respectively.
- Monthly quality control includes quantifying and identifying a viable culture to genus level.
- Prior to analyzing samples, each microscope's Kohler illumination is checked. The microscope fields of view are calibrated annually.
- The lactophenol dye, slides, cover slips and spore traps are checked on a daily basis to assure that there is no contamination. Upon initial receipt, one spore trap from each lot that SGS Galson receives is checked for possible contamination.
- Media used for viable analysis is tested upon receipt for both sterility and growth promotion.
- A second analyst reexamines samples that have no observable spores.
- All reports undergo a secondary quality assurance review prior to release.

L615648

16

GALSON CHAIN OF CUSTODY

You may edit and complete this COC electronically by logging in to your Client Portal account at <https://portal.galsonlabs.com/>

Turn Around Time (TAT): (surcharge)	<input checked="" type="checkbox"/> Standard 0% <input type="checkbox"/> 4 Business Days 35% <input type="checkbox"/> 3 Business Days 50% <input type="checkbox"/> 2 Business Days 75% <input type="checkbox"/> Next Day by 6pm 100% <input type="checkbox"/> Next Day by Noon 150% <input type="checkbox"/> Same Day 200%	Client Acct No.: <u>13111</u> Report To: <u>Clint Smith</u> Company Name: <u>Colden Corporation</u> Address 1: <u>5842 Heritage Landing Dr.</u> Address 2: _____ City, State Zip: <u>East Syracuse, NY 13057</u> Phone No.: <u>315 - 445 - 0847</u> Cell No.: <u>315 - 404 - 8075</u> Email reports to: <u>smith@colden.com</u> Email EDD to: <u>smith@colden.com</u> Comments: _____	Invoice To: <u>Accounts Payable</u> Company Name: <u>Colden Corporation</u> Address 1: <u>5842 Heritage Landing Dr.</u> Address 2: _____ City, State Zip: <u>East Syracuse, NY 13057</u> Phone No.: <u>315 - 445 - 0847</u> Email Address: <u>admin@colden.com</u> Comments: _____ P.O. No.: _____ Payment info.: <input type="checkbox"/> I will call SGS Galson to provide credit card info <input type="checkbox"/> Card on File (enter the last five digits on the line below)
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Original Prep No.: PSY725760
 CS Rep: JTRAINER
 Online COC No.: 288362

Samples submitted using the FreePumpLoan™ Program
 Samples submitted using the FreeSamplingBadges™ Program

Comments: _____

State Sampled: NY

Please indicate which OEL(s) this data will be used for:
 OSHA PEL ACGIH TLV MSHA Cal OSHA
 MLO: n/a Other: _____
 Specify Limit(s) Specify Other

Site Name: HC Project: 24123 Sampled By: C. Smith

List description of industry or Process/interferences present in sampling area: _____

Sample ID * (Maximum of 20 Characters)	Date Sampled *	Collection Medium	Sample Volume Sample Time Sample Area *	Liters Minutes in ² , cm ² , ft ² *	Analysis Requested	Method Reference ^	Hexavalent Chromium Process (e.g., welding, plating, painting, etc.)
<u>HC240119-001</u>	<u>1/19/24</u>	<u>CultureSwab</u>	<u>n/a</u>	<u>n/a</u>	<u>Standard Mold Screen</u>	<u>In-house: MICR-SOP-21; Microscopy</u>	
<u>↓ -002</u>	<u>↓</u>	<u>CultureSwab</u>	<u>↓</u>	<u>↓</u>	<u>Standard Mold Screen</u>	<u>In-house: MICR-SOP-21; Microscopy</u>	

^ If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature	Date	Time	Received By:	Date	Time
Relinquished By:	<u>Clint Smith</u>	<u>1/19/24</u>	<u>11:45</u>	<u>JOSHUA PENALS</u>	<u>1/19/24</u>	<u>11:45</u>
Relinquished By:	<u>JOSHUA PENALS</u>	<u>1/19/24</u>	<u>13:50</u>	<u>Kathryn L. Drogo</u>	<u>1/19/24</u>	<u>14:02</u>

Online COC No.: 288362
 Prep No.: PSY725760
 Account No.: 13111
 Draft: 1/17/2024 7:54:32 AM

* You must fill in these columns for any samples which you are submitting.
 Samples received after 3pm will be considered as next day's business.

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>



GALSON

CHAIN OF CUSTODY

Comments :

Sample ID * (Maximum of 20 Characters)	Date Sampled *	Collection Medium	Sample Volume Sample Time Sample Area *	Liters Minutes in ² , cm ² , ft ² *	Analysis Requested	Method Reference ^	Hexavalent Chromium Process (e.g., welding, plating, painting, etc.)
HC240119-003	1/19/24	Culture Swab	n/a	n/a	Standard Mold Screen	In-house: MICR-SOP-21; Microscopy	
-004		Air-O-Cell culture Swab			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-005		Air-O-Cell culture Swab			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-006		Air-O-Cell culture Swab			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-007		Air-O-Cell culture Swab			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-008A		Air-O-Cell	14.98		Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-008B		Air-O-Cell	29.96		Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-009		Air-O-Cell culture Swab	n/a	n/a	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	

^ If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature		Date	Time	Print Name / Signature		Date	Time
Relinquished By:	Clint Smith	<i>Clint Smith</i>	1/19/24	11:45	Received By:	JOSHUA PINALS	1/19/24	11:45
Relinquished By:	JOSHUA PINALS	<i>Joshua Pinals</i>	1/19/24	13:50	Received By:	Kathryn L. Drogo	1/19/24	14:02

* You must fill in these columns for any samples which you are submitting. Samples received after 3pm will be considered as next day's business.

Online COC No. : 288362
Prep No. : PSY725760
Account No. : 13111
Draft : 1/17/2024 7:54:32 AM

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>



GALSON

Clint Smith
Colden Corporation
5842 Heritage Landing Dr.
East Syracuse, NY 13057

January 31, 2024

Account# 13111

Login# L616462

Dear Clint Smith:

Enclosed are the analytical results for the samples received by our laboratory on January 31, 2024. All samples on the chain of custody were received in good condition unless otherwise noted. Any additional observations will be noted on the chain of custody.

Please contact client services at (888) 432-5227 if you would like any additional information regarding this report. Thank you for using SGS Galson.

Sincerely,

SGS Galson

Lisa Swab
Laboratory Director

Enclosure(s)



Terms and Conditions & General Disclaimers

- This document is issued by the Company under its General Conditions of Service accessible at <http://www.sgs.com/en/Terms-and-Conditions.aspx>. Attention is drawn to the limitation of liability, indemnification and jurisdiction issues defined therein.
- Any holder of this document is advised that information contained herein reflects the Company’s findings at the time of its intervention only and within the limits of Client’s instructions, if any. The Company’s sole responsibility is to its Client and this document does not exonerate parties to a transaction from exercising all their rights and obligations under the transaction documents. Any unauthorized alteration, forgery or falsification of the content or appearance of this document is unlawful and offenders may be prosecuted to the fullest extent of the law.

Analytical Disclaimers

- Unless otherwise noted within the report, all quality control results associated with the samples were within established control limits or did not impact reported results.
- Note: The findings recorded within this report were drawn from analysis of the sample(s) provided to the laboratory by the Client (or a third party acting at the Client’s direction). The laboratory does not have control over the sampling process, including but not limited to the use of field equipment and collection media, as well as the sampling duration, collection volume or any other collection parameter used by the Client. The findings herein constitute no warranty of the sample's representativeness of any sampled environment, and strictly relate to the samples as they were presented to the laboratory. For recommended sampling collection parameters, please refer to the Sampling and Analysis Guide at www.sgsgalson.com.
- Unrounded results are carried through the calculations that yield the final result and the final result is rounded to the number of significant figures appropriate to the accuracy of the analytical method. Please note that results appearing in the columns preceding the final result column may have been rounded and therefore, if carried through the calculations, may not yield an identical final result to the one reported.
- The stated LOQs for each analyte represent the demonstrated LOQ concentrations prior to correction for desorption efficiency (if applicable).
- Unless otherwise noted within the report, results have not been blank corrected for any field blank or method blank data.

Accreditations SGS Galson holds a variety of accreditations and recognitions. Our quality management system conforms with the requirements of ISO/IEC 17025. Where applicable, samples may also be analyzed in accordance with the requirements of ELAP, NELAC, or LELAP under one of the state accrediting bodies listed below. Current Scopes of Accreditation can be viewed at <http://www.sgsgalson.com> in the accreditations section of the "About" page. To determine if the analyte tested falls under our scope of accreditation, please visit our website or call Client Services at (888) 432-5227.

National/International	Accreditation/Recognition	Lab ID#	Program/Sector
AIHA-LAP, LLC - IHLAP, ELLAP, EMLAP	ISO/IEC 17025 and USEPA NLLAP	Lab ID 100324	Industrial Hygiene, Environmental Lead, Environmental Microbiology

State	Accreditation/Recognition	Lab ID#	Program/Sector
New York (NYSDOH)	ELAP and NELAC (TNI)	Lab ID: 11626	Air Analysis, Solid and Hazardous Waste
Louisiana (LDEQ)	LELAP	Lab ID: 04083	Air Analysis, Solid Chemical Materials

Legend

< - Less than	mg - Milligrams	MDL - Method Detection Limit	ppb - Parts per Billion
> - Greater than	ug - Micrograms	NA - Not Applicable	ppm - Parts per Million
l - Liters	m3 - Cubic Meters	NS - Not Specified	ppbv - ppb Volume
LOQ - Limit of Quantitation	kg - Kilograms	ND - Not Detected	ppmv - ppm Volume
ft2 - Square Feet	cm2 - Square Centimeters	in2 - Square Inches	ng - Nanograms



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 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A125B Lab ID : L616462-2 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	1	1	13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	17	17	230	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	11	11	150	64.7
Basidiospores	5	5	67	29.4
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	1	1	13	5.9
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A126A Lab ID : L616462-3 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	1	1	13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	437	437	5800	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	430	430	5700	98.4
Basidiospores	4	4	53	0.9
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	3	3	40	0.7
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A126B Lab ID : L616462-4 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	1	1	13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	342	562	7500	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	340	560	7500	99.6
Basidiospores	2	2	27	0.4
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A120A Lab ID : L616462-5 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	12	12	160	NA
Pollen	1	1	13	NA
Total Fungal Spores	257	257	3400	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	3	3	40	1.2
<i>Aspergillus/Penicillium</i> -like	231	231	3100	89.9
Basidiospores	12	12	160	4.7
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	4	4	53	1.6
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	2	2	27	0.8
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	5	5	67	1.9

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A120B Lab ID : L616462-6 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	11	11	150	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	514	784	10000	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	11	11	150	1.4
<i>Aspergillus/Penicillium</i> -like	420	690	9200	88
Basidiospores	12	12	160	1.5
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	67	67	890	8.5
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	4	4	53	0.5

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A119A Lab ID : L616462-7 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	3	3	40	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	53	53	710	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	34	34	450	64.2
Basidiospores	5	5	67	9.4
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	13	13	170	24.5
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	1.9

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A119B Lab ID : L616462-8 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	4	4	53	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	339	339	4500	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	2	2	27	0.6
<i>Aspergillus/Penicillium</i> -like	203	203	2700	59.9
Basidiospores	21	21	280	6.2
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	107	107	1400	31.6
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	4	4	53	1.2
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	2	2	27	0.6

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B125A Lab ID : L616462-9 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	401	1201	16000	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	400	1200	16000	99.9
Basidiospores	1	1	13	0.1
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

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 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B125B Lab ID : L616462-10 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	1	1	13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	119	119	1600	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	117	117	1600	98.3
Basidiospores	<1	<1	<13	NA
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	2	2	27	1.7
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B119A Lab ID : L616462-11 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	40	40	530	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	18	18	240	45
Basidiospores	22	22	290	55
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

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 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B119B Lab ID : L616462-12 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	2	2	27	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	60	60	800	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	44	44	590	73.3
Basidiospores	13	13	170	21.7
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	3	3	40	5
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B109A Lab ID : L616462-13 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	1	1	13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	339	659	8800	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium-like</i>	320	640	8500	97.1
Basidiospores	6	6	80	0.9
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	5	5	67	0.8
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	8	8	110	1.2

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B109B Lab ID : L616462-14 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	2	2	27	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	362	978	13000	NA

<i>Alternaria</i>	1	1	13	0.1
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	344	960	13000	98.2
Basidiospores	3	3	40	0.3
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	13	13	170	1.3
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	0.1

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 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B107A Lab ID : L616462-15 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 1

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	1	1	13	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium-like</i>	<1	<1	<13	NA
Basidiospores	1	1	13	100
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

CFU -Colony Forming Units g -Grams



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LABORATORY ANALYSIS REPORT

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 East Syracuse, NY 13057
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 FAX: (315) 437-0571
 www.sgsgalson.com

Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-B107B Lab ID : L616462-16 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	13	13	170	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium-like</i>	9	9	120	69.2
Basidiospores	2	2	27	15.4
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	2	2	27	15.4

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

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Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A105A Lab ID : L616462-17 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	16	16	210	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	9	9	120	56.3
Basidiospores	1	1	13	6.3
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	5	5	67	31.3
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	6.3

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

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 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A105B Lab ID : L616462-18 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	1	1	13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	16	16	210	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	2	2	27	12.5
<i>Aspergillus/Penicillium</i> -like	8	8	110	50
Basidiospores	2	2	27	12.5
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	1	1	13	6.3
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	3	3	40	18.8

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

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Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A118A Lab ID : L616462-19 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	62	62	830	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	61	61	810	98.4
Basidiospores	1	1	13	1.6
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

CFU -Colony Forming Units g -Grams



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Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-A118B Lab ID : L616462-20 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 1

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	3	3	40	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	1	1	13	33.3
Basidiospores	1	1	13	33.3
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	33.3

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

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Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-OUT1 Lab ID : L616462-21 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 1

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	<1	<1	<13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	2	2	27	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	<1	<1	<13	NA
Basidiospores	<1	<1	<13	NA
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	<1	<1	<13	NA
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	2	2	27	100

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

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 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-OUT2 Lab ID : L616462-22 Air Volume : 0.075 m3
 Analysis : Standard Mold Screen Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	1	1	13	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	30	30	400	NA

<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	3.3
<i>Aspergillus/Penicillium</i> -like	1	1	13	3.3
Basidiospores	16	16	210	53.3
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	7	7	93	23.3
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	5	5	67	16.7

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

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Client : Colden Corporation
Site : HC
Project No. : 24123
Date Sampled : 30-JAN-24
Date Received : 31-JAN-24
Incubation Temp : NA

Account No.: 13111
Login No. : L616462
Date Analyzed : 31-JAN-24
Report ID : 1404712

Client ID : HC240130-BL1 Lab ID : L616462-23 Air Volume : NA
Analysis : Standard Mold Screen Crowding Factor : 0

Table with 4 columns: Parameter, Raw Count, Total Count, Percent %. Rows include Mycelial Fragments, Pollen, Total Fungal Spores, Alternaria, Ascospores, Aspergillus/Penicillium-like, Basidiospores, Bipolaris/Drechslera, Chaetomium, Cladosporium, Curvularia, Rusts/Smuts, Stachybotrys, and Other/Unidentified.

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

CFU -Colony Forming Units g -Grams



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LABORATORY ANALYSIS REPORT

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Client : Colden Corporation
 Site : HC
 Project No. : 24123
 Date Sampled : 30-JAN-24
 Date Received : 31-JAN-24
 Incubation Temp : NA

Account No.: 13111
 Login No. : L616462
 Date Analyzed : 31-JAN-24
 Report ID : 1404712

Client ID : HC240130-BL2 Lab ID : L616462-24 Air Volume : NA
 Analysis : Standard Mold Screen Crowding Factor : 0

Parameter	Raw Count	Total Count	Percent %
Mycelial Fragments	<1	<1	NA
Pollen	<1	<1	NA
Total Fungal Spores	<1	<1	NA

<i>Alternaria</i>	<1	<1	NA
Ascospores	<1	<1	NA
<i>Aspergillus/Penicillium</i> -like	<1	<1	NA
Basidiospores	<1	<1	NA
<i>Bipolaris/Drechslera</i>	<1	<1	NA
<i>Chaetomium</i>	<1	<1	NA
<i>Cladosporium</i>	<1	<1	NA
<i>Curvularia</i>	<1	<1	NA
Rusts/Smuts	<1	<1	NA
<i>Stachybotrys</i>	<1	<1	NA
Other/Unidentified	<1	<1	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore Submitted by: SLS/TAC Supervisor: BDB Date : 31-JAN-24
 Analytical Method : In-house: IB-AIROCELL; Mic Approved by : BDB Sampler : Spore Trap

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LABORATORY FOOTNOTE REPORT

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Client Name : Colden Corporation
Site : HC
Project No. : 24123

Date Sampled : 30-JAN-24
Date Received: 31-JAN-24
Date Analyzed: 31-JAN-24

Account No.: 13111
Login No. : L616462

L616462 (Report ID: 1404712):
SOPs: ib-airocell(29)



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 Fax: (315) 437-0571
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Analytical Notes for Microbiology Air-O-Cell™ Cassettes and other Spore Traps

Air-O-Cell™ cassettes and other spore traps may capture non-microbial particles that may interfere with spore counts. SGS Galson provides an estimation of the density of these particles, referred to as a Crowding Factor. The Crowding Factor ranges from 0 to 5 and is explained below. High levels of particulate matter on the impaction medium may bias the analysis by obscuring or covering spores. In addition, particle capture efficiency may decrease with high levels of particulate matter.

Crowding Factor	Explanation
0	No particles detected. This is typical of blank samples. Because most air samples typically contain some particles, absence of particulate matter could indicate improper sampling if the sample was not meant to be a blank.
1	Particles are far apart and in low numbers. Particulate matter covers approximately <5% of the impaction area. Spore counts not affected or minimally affected by the particle load.
2	Particles are close together and/or overlapping, and some spores may be obscured. Particulate matter covers approximately 5% to 25% of the impaction area. Spore counts may be biased low.
3	Particles are moderately crowded. It is likely that some spores are obscured. Particulate matter covers approximately 25% to 75% of the impaction area. Spore counts are likely biased low.
4	Particles are crowded, frequently obscuring spores. Particulate matter covers approximately 75% to 90% of the impaction area. Spore counts are likely biased low. The degree of bias increases with the percent of the trace that is occluded.
5	Particles are overcrowded making analysis impossible; no spore counts provided. If certain spores are readily detectable, they are reported as "Detected". If heavy quantities of spores are observed along the edges of the trace, this is footnoted in the report.



Counts for any genus that exceed 300 spores are estimated to two significant figures.

The list of fungal spores reported is:

***Alternaria* includes spores previously reported as *Ulocladium*.**

Ascospores – includes all ascospores with the exception of *Chaetomium*.

***Aspergillus/Penicillium*-like** – These two genera are grouped together as the spores are indistinguishable on a spore trap.

Basidiospores – This includes all basidiospores, even ones that can be identified to genus level, such as *Ganoderma*.

Bipolaris/Drechslera – *Helminthosporium* and *Exserohilum* are included in this grouping.

Chaetomium – Due to its unique shape and due to the fact that it may be associated with indoor mold problems, this ascospore is reported separate from other ascospores.

Cladosporium

Curvularia

Rusts/Smuts – *Myxomycetes* and *Periconia* are included in this grouping.

Stachybotrys – This includes *Memnoniella*.

Ulocladium* has been reclassified and is now reported as *Alternaria

Other/Unidentified – “Other” includes spores that can be identified but are rarely observed and/or are typically seen in small quantities. They include: *Acremonium*, *Botrytis*, *Cercospora*, *Epicoccum*, *Fusarium*, *Nigrospora*, *Oidium*, *Paecilomyces*, *Pestalotia*, *Pestalotiopsis*, *Pithomyces*, *Polythrincium*, *Scopulariopsis*, *Spegazzinia*, *Stemphylium*, *Taeniocella*, *Tetraploa*, *Torula*, and *Trichoderma*, and *Zygophiala*. “Unidentified” includes broken and dehydrated spores, spores that are partially obscured by debris, and spores that can't be categorized using microscopy alone.

In addition, other analytes that will be shown on reports include mycelial fragments (hyphae) and pollen.

Reports for expanded analysis include the above list with the addition of skin cells and fibers.

Generally, 100% of the sample deposit is analyzed. However, some analytes with high counts may be estimated based on the analysis of a portion of the slide and the results extrapolated. In these cases, the reported values will differ between the “Raw Count” and “Total Count” columns. For example, if an analyst observed 304 basidiospores after analyzing 25% of the sample, the estimated value is 1216. The final report would show 304 in the “Raw Count” column and 1200 in the “Total Count” column (the “Total Column” is rounded to two significant figures).



Direct Microscopic Examination (Screens)

- The analytes that we report are the same as those listed for spore traps with the exceptions of pollen, skin cells, and fibers.
- Due to the inherent nature of screen samples, a spore count is not performed.
- Upon special request counts may be performed on swab, liquid, or bulk screens. Counts are never performed on tape lifts due to the nature of the samples to not have uniform distribution of spores.
- The amount of a particular spore detected is reported as a “Level of contamination”. The level of contamination is a subjective measurement and corresponds to the general quantity of spores present in a sample. It also describes the amount of spores relative to one another.
 - Light: approximately 1 to 5 spores or mycelial fragments per microscope field of view at 600x.
 - Moderate: 6 to 15 spores or mycelial fragments per microscope field of view at 600x.
 - Heavy: Greater than 15 spores or mycelial fragments per microscope field of view at 600x.

Viable Fungi Analysis

- Standard growing conditions for viable fungi are $25^{\circ}\text{C} \pm 1^{\circ}\text{C}$ for 7 days.
- Standard growing conditions for viable thermophilic fungi are $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$ for 7 days.
- Results are reported in colony forming units (CFUs). A CFU can originate from one or many spores.
- SGS Galson uses and provides Potato Dextrose agar for all cultureable fungal methods. We have found Potato Dextrose agar to be suitable for the culture of the widest range of organisms. Other agars submitted or requested by clients are grown under the above standard conditions unless otherwise requested by the client.
- Some fungi may not produce identifiable structures in culture or under standard growing conditions. These fungi will be considered sterile hyphae and reported as such.
- Lack of growth under standard conditions does not preclude the presence of fungi or its viability in a sample.
- Samples taken with impactor samplers are not corrected for a positive hole correction factor.
- Identification of fungal organisms is based on visual microscopic examination at up to seven days of growth under standard conditions. Due to the large numbers of different species that may comprise them, certain genera may appear similar due to variations in stages of their life cycles, growth requirements, and/or environmental stress. A very limited amount of identification overlap may occur due to morphological similarities.
- Final interpretation of results is up to the person(s) responsible for conducting the sampling.



Quality Control/Quality Assurance

- A daily quality control spore trap slide is read each day that an analyst performs analysis on client spore trap samples. These slides consist of old client samples that have been analyzed a minimum of twenty times before they are used as a part of the quality control program. Control limits are set at the mean plus or minus three standard deviations for each analyte and for the total spore count. Warning limits are set at the mean plus or minus two standard deviations for each analyte and for the total spore count.
- A minimum of five percent of the samples are analyzed as duplicates and five percent of the samples are analyzed as replicates (or at least one replicate or duplicate per day). The relative percent difference (RPD) is calculated between the original sample result and its duplicate or replicate. The RPD value must fall within statistically based limits. In addition, there must be agreement between three of the top five categories.
- Daily quality control includes a blind spore trap challenge and a blind fungal culture identification challenge. Each analyst must correctly identify a spore or other airborne particulate from an old spore trap slide and identify a slide prepared from a fungal culture, respectively.
- Monthly quality control includes quantifying and identifying a viable culture to genus level.
- Prior to analyzing samples, each microscope's Kohler illumination is checked. The microscope fields of view are calibrated annually.
- The lactophenol dye, slides, cover slips and spore traps are checked on a daily basis to assure that there is no contamination. Upon initial receipt, one spore trap from each lot that SGS Galson receives is checked for possible contamination.
- Media used for viable analysis is tested upon receipt for both sterility and growth promotion.
- A second analyst reexamines samples that have no observable spores.
- All reports undergo a secondary quality assurance review prior to release.

COLDEN_CORP
 Date: 01/31/24
 Shipper: DROP OFF
 Initials: MMM



Prep: UNKNOWN

Handwritten: L16647 ^{KMS} L166462
GALSON

CHAIN OF CUSTODY

Handwritten: R125

may edit and complete this COC electronically by logging in to your Client Portal account at <https://portal.galsonlabs.com/>

<input type="checkbox"/>	Standard	0%
<input type="checkbox"/>	4 Business Days	35%
<input type="checkbox"/>	3 Business Days	50%
<input type="checkbox"/>	2 Business Days	75%
<input type="checkbox"/>	Next Day by 6pm	100%
<input type="checkbox"/>	Next Day by Noon	150%
<input checked="" type="checkbox"/>	Same Day	200%
<input type="checkbox"/>	Samples submitted using the FreePumpLoan™ Program	
<input type="checkbox"/>	Samples submitted using the FreeSamplingBadges™ Program	

Client Acct No.: 13111	Report To: Clint Smith	Invoice To: Accounts Payable
Company Name: Colden Corporation	Company Name: Colden Corporation	Company Name: Colden Corporation
Address 1: 5842 Heritage Landing Dr.	Address 1: 5842 Heritage Landing Dr.	Address 1: 5842 Heritage Landing Dr.
Address 2:	Address 2:	Address 2:
Original Prep No.: PSY727118	City, State Zip: East Syracuse, NY 13057	City, State Zip: East Syracuse, NY 13057
CS Rep: JTRAINER	Phone No.: 315 - 445 - 0847	Phone No.: 315 - 445 - 0847
Online COC No.: 289115	Cell No.: 315 - 404 - 8075	Email Address: admin@colden.com
	Email reports to: smith@colden.com	Comments:
	Email EDD to: smith@colden.com ; magari@colden.com	P.O. No.:
	Comments: shepard@colden.com	Payment info.: <input type="checkbox"/> I will call SGS Galson to provide credit card info <input type="checkbox"/> Card on File (enter the last five digits on the line below)

Comments:

State Sampled: **NY**

Please indicate which OEL(s) this data will be used for:
 OSHA PEL ACGIH TLV MSHA Cal OSHA
 IAQ: **n/a** Other: _____
 Specify Limit(s) Specify Other

Site Name: **HC** Project: **24123** Sampled By: **C. Smith**

List description of industry or Process/interferences present in sampling area:

Sample ID * (Maximum of 20 Characters)	Date Sampled *	Collection Medium	Sample Volume Sample Time Sample Area *	Liters Minutes in ² , cm ² , ft ² *	Analysis Requested	Method Reference ^	Hexavalent Chromium Process (e.g., welding, plating, painting, etc.)
HC240130-A125A	1/30/24	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
↓ - A125B	↓	Air-O-Cell	↓	↓	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	

^ If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature	Date	Time	Print Name / Signature	Date	Time
Relinquished By: Clint Smith	<i>[Signature]</i>	1/31/24	0930	Received By: Megan M. McGrath	1/31/24	9:45
Relinquished By:				Received By:		

*** You must fill in these columns for any samples which you are submitting.**

Samples received after 3pm will be considered as next day's business.

Online COC No.: **289115**
 Prep No.: **PSY727118**
 Account No.: **13111**
 Draft: **1/29/2024 3:05:05 PM**

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>



GALSON

CHAIN OF CUSTODY

Comments :

Sample ID * (Maximum of 20 Characters)	Date Sampled *	Collection Medium	Sample Volume Sample Time Sample Area *	Liters Minutes in ² , cm ² , ft ² *	Analysis Requested	Method Reference ^	Hexavalent Chromium Process (e.g., welding, plating, painting, etc.)
HC240130 - A126A	1/30/24	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-A126B		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-A120A		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-A120B		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-A119A		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-A119B		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-B125A		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-B125B		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-B119A		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-B119B		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
-B109A		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	

^ If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature		Date	Time	Print Name / Signature		Date	Time
Relinquished By:	Clat Smith	<i>Clat Smith</i>	1/31/24	0930	Received By:	Megan M. McGrath	1/31/24	9:45
Relinquished By:					Received By:			

* You must fill in these columns for any samples which you are submitting.

Samples received after 3pm will be considered as next day's business.

Online COC No. : 289115
Prep No. : PSY727118
Account No. : 13111
Draft : 1/29/2024 3:05:05 PM

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>

Comments :

Sample ID * (Maximum of 20 Characters)	Date Sampled *	Collection Medium	Sample Volume Sample Time Sample Area *	Liters Minutes in ² , cm ² , ft ² *	Analysis Requested	Method Reference ^	Hexavalent Chromium Process (e.g., welding, plating, painting, etc.)
HC240130 - B107B	1/30/24	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
- B107A		Air-O-Cell			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	
- B107B							
- A105A							
- A105B							
- A118A							
- A118B							
- OUT1							
- OUT2							
HC240130 - BL1	1/30/24	Air-O-cell	BLANK →				
- BL2			BLANK →				

^ If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature	Date	Time	Print Name / Signature	Date	Time
Relinquished By:	C. A. Smith	1/30/24	0930	Received By: Megan M. McGrath	1/31/24	9:45
Relinquished By:				Received By:		

* You must fill in these columns for any samples which you are submitting.

Samples received after 3pm will be considered as next day's business.

Online COC No. : 289115
 Prep No. : PSY727118
 Account No. : 13111
 Draft : 1/29/2024 3:05:05 PM

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>

ATTACHMENT C
Equipment Calibration Certificates

1st Round of Sampling



This should NOT be used as a Chain of Custody

Laboratory Pump Calibration Data

Prep #:

Pump Calibration Record:

RSY725760

Page 1

Date: 01/17/24	If Aircheck Battery Number	Target Flow Rate	Calibrated by: JAH (initials)	Type of Media and/or Method	Post-Calibrated By: (initials and date)	Average of Pre- and Post- readings
Pump Number			Pre-Calibration Flow Rate		Laboratory Post-Calibration Flow Rate *	
BP053		15.00	14.98	AOC		

Primary Calibrator: MFM002 Date: _____

Box checked if
calibrated with
a Dry-Cal.

All pumps are calibrated with a TSI Primary Calibrator unless otherwise specified.

Postcal: Place the red Post Calibration Required sticker on any pumps that require Laboratory Post Calibration. Post calibrations performed by SGS Galson will be utilized if either a) post-calibration is outside 5% of pre-calibration or b) average of pre- and post-calibration flow rates is outside of 5% of target flow rate. (For flow rates below 0.1 L/min, we will use 10% criteria due to the limitation of calibrator precision.) Sample-pump correlations must be provided by client in order to apply post-calibration. Pumps post calibrated by SGS Galson are not performed with the sampling matrix in line and may not account for loss of pressure during sampling.

* Laboratory post-calibration is not recommended. Field calibrations are preferred as they account for site conditions which may affect flow rates.

*2nd Round of Sampling
Jan. 30, 2024*



This should NOT be used as a Chain of Custody

Laboratory Pump Calibration Data

Prep #:

Pump Calibration Record:

RSY727118

Page 1

Date: 01/29/24		Target Flow Rate	Calibrated by: MJC (initials)		Type of Media and/or Method	Post-Calibrated By: (Initials and date)		Average of Pre- and Post- readings
Pump Number	If Aircheck Battery Number		Pre-Calibration Flow Rate			Laboratory Post-Calibration Flow Rate*		
BP034		15.00	14.99		AOC			

Primary Calibrator: MFM001 Date: 12/31/23

Box checked if calibrated with a Dry-Cal.

All pumps are calibrated with a TSI Primary Calibrator unless otherwise specified.

Postal: Place the red Post Calibration Required sticker on any pumps that require Laboratory Post Calibration. Post calibrations performed by SGS Galson will be utilized if either a) post-calibration is outside 5% of pre-calibration or b) average of pre- and post-calibration flow rates is outside of 5% of target flow rate. (For flow rates below 0.1 L/min, we will use 10% criteria due to the limitation of calibrator precision.) Sample-pump correlations must be provided by client in order to apply post-calibration. Pumps post calibrated by SGS Galson are not performed with the sampling matrix in line and may not account for loss of pressure during sampling.

* Laboratory post-calibration is not recommended. Field calibrations are preferred as they account for site conditions which may affect flow rates.